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<b>(21) International Application Number:</b> PCT/GB99/01059 <b>(22) International Filing Date:</b> 7 April 1999 (07.04.99) <b>(30) Priority Data:</b> 9807818.1      9 April 1998 (09.04.98)      GB <b>(71) Applicant (for all designated States except US):</b> ZENECA LIMITED [GB/GB]; 15 Stanhope Gate, London W1Y 6LN (GB). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> SHIPTON, Catherine, Ann [GB/GB]; Jealotts Hill Research Station, Bracknell, Berkshire RG42 6ET (GB). BRYAN, Ian, Bennett [GB/GB]; Jealotts Hill Research Station, Bracknell, Berkshire RG42 6ET (GB). <b>(74) Agents:</b> GAAL, Jozsef, Christopher et al.; Zeneca Agrochemicals, Intellectual Property Dept., P.O. Box 3538, Jealott's Hill Research Station, Bracknell, Berkshire RG42 6YA (GB).		<b>(81) Designated States:</b> AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
<b>(54) Title:</b> METHOD OF PRODUCING PLANTS WHICH ARE TOLERANT OR RESISTANT TO HERBICIDES		
<b>(57) Abstract</b>  A method of making plants which are resistant or tolerant to herbicides which, <i>in vitro</i> , inhibit 4-hydroxyphenylpyruvate dioxygenase (4HPPD) comprises the steps of: (i) transforming plant material with a polynucleotide comprising a region encoding a phytoene desaturase; (ii) regenerating the thus transformed material into morphologically normal plants. In a preferred embodiment the region comprised by the polynucleotide is the sequence depicted in SEQ ID No.1, or is a sequence which is complementary to one which when incubated at a temperature of between 55 and 60 °C in 0.3 strength citrate buffered saline containing 0.1 % SDS followed by rinsing at the same temperature with 0.3 strength citrate buffered saline containing 0.1 % SDS still hybridises with the sequence depicted in SEQ ID No.1.		

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METHOD OF PRODUCING PLANTS WHICH ARE TOLERANT OR RESISTANT TO  
HERBICIDES

The present invention relates *inter alia*, to a method of producing plants which are  
5 tolerant or resistant to herbicides and in particular to the production of transgenic plants  
which exhibit substantial resistance or substantial tolerance to herbicides when compared  
with non transgenic like plants.

Plants which are substantially "tolerant" to a herbicide when they are subjected to it provide  
a dose/response curve which is shifted to the right when compared with that provided by  
10 similarly subjected non tolerant like plants. Such dose/response curves have "dose" plotted  
on the x-axis and "percentage kill", "herbicidal effect" etc. plotted on the y-axis. Tolerant  
plants will require more herbicide than non tolerant like plants in order to produce a given  
herbicidal effect. Plants which are substantially "resistant" to the herbicide exhibit few, if  
any, necrotic, lytic, chlorotic or other lesions when subjected to the herbicide at  
15 concentrations and rates which are typically employed by the agrochemical community to  
kill weeds in the field. Plants which are resistant to a herbicide are also tolerant of the  
herbicide. The terms "resistant" and "tolerant" are to be construed as "tolerant and/or  
resistant" within the context of the present application.

The herbicides of particular relevance to the present invention are those which are  
20 capable *in vitro* of inhibiting 4-Hydroxy-phenylpyruvate dioxygenase (HPPD or 4HPPD)  
enzymes. Such herbicides have been disclosed, such as the isoxazoles described especially in  
the French Patent Applications 95 06800 and 95 13570 and especially isoxaflutole, a  
selective maize herbicide, diketonitriles such as those described in European Applications 0  
496 630, 0496 631, in particular 2-cyano-3-cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-CF<sub>3</sub>-  
25 phenyl)propane-1,3-dione and 2-cyano-3-cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-2,3Cl<sub>2</sub>phenyl)propane-  
1,3-dione, triketones described in European Applications 0 625 505 and 0 625 508, in  
particular sulcotrione, mesotrione (BSI-proposed), pyrazolynate and pyrazoxyfen. Known  
genes capable of providing for tolerance to these herbicides are those which encode HPPD  
enzymes.

According to the present invention there is provided a method of making plants which are resistant or tolerant to herbicides which - *in vitro* - inhibit 4-hydroxyphenylpyruvate dioxygenase (4HPPD) comprising the steps of:

- (i) transforming plant material with a polynucleotide comprising a region encoding a phytoene desaturase (PDS);
- (ii) regenerating the thus transformed material into morphologically normal plants.

The region comprised by the polynucleotide may have the sequence depicted in SEQ ID No. 1, or may be a sequence which is complementary to one which when incubated at a temperature of between 55 and 60°C in 0.3 strength citrate buffered saline containing 0.1% SDS followed by rinsing at the same temperature with 0.3 strength citrate buffered saline containing 0.1% SDS still hybridises with the sequence depicted in SEQ ID No. 1.

It is preferred that the phytoene desaturase is of bacterial origin such as that depicted in SEQ ID No. 1 and being derived from *Erwinia uredovora*, and/or in particular is one which does not require plastoquinone 9 as a co-factor. The desaturase may, however be of plant origin, such as especially of monocotyledonous or dicotyledonous plants, especially of *Arabidopsis* or of *Umbelliferae*, such as, for example, the carrot (*Daucus carotta*). It can be native or possibly mutated while at the same time fundamentally retaining a property of herbicidal tolerance against HPPD inhibitors, such as herbicides of the isoxazoles family such as the Balance™ Herbicide or triketones. The herbicide resistant plants produced by the above method may be selected through their resistance to herbicides which *in vitro*, inhibit 4HPPD.

It may however, be further preferred that the polynucleotide encoding the phytoene desaturase further comprises a selectable marker gene to facilitate the selection of regenerated transformants. Suitable selectable marker genes include; resistance to antibiotics such as kanamycin, hygromycin and gentamycin; resistance to further herbicides such as glyphosate based herbicides; resistance to toxins such as eutypine.

Other forms of selection are also available such as hormone based selection systems such as the Multi Auto Transformation (MAT) system of Hiroyasu Ebinuma *et al.* 1997. PNAS Vol. 94 pp2117-2121; visual selection systems which use the known green fluorescence protein,  $\beta$  glucuronidase, mannose isomerase, xylose isomerase and 2-DOG.

The plant material may be, or may have been, further transformed with a polynucleotide comprising a region encoding a protein capable of providing the plant material with

resistance or tolerance to herbicides, insects, desiccation and/or fungal, bacterial or viral infections, or with a polynucleotide capable of encoding proteins which provide for improved quality traits such as increased yield, altered starch quality and/or increased nutrient content.

5           The protein encoding sequences within the polynucleotide are bounded by plant operable promoters and terminators. Such promoters and terminators, which are *per se* not germane to the invention, are well known to the skilled man and include, for example, the CaMV35S, FMV35S, NOS, OCS and E9 (derived from the small subunit of RUBISCO) promoters and terminators, or the promoter and terminator of a gene of alpha-tubulin (EP-A  
10 652,286). Preferably, recourse is made to a promoter regulation sequence which favours the over-expression of the coding sequence, such as, for example, that comprising at least one histone promoter such as described in EP-A-507,698.

          According to the invention, it is equally possible to use, in association with the promoter regulation sequence, other regulation sequences which are situated between the  
15 promoter and the coding sequence, such as transcriptional or translational enhancers such as, for example, tobacco etch virus (TEV) translation activator described in International Patent application, PCT publication number WO87/07644 which is incorporated herein by reference, or of transit peptides, either single, or double, and in this case possibly separated by an intermediate sequence, that is to say comprising, in the transcription direction, a  
20 sequence coding for a transit peptide of a plant gene coding for a plastid localization enzyme, a part of the sequence of the N-terminal mature part of a plant gene coding for a plastid localization enzyme, then a sequence coding for a second transit peptide of a plant gene coding for a plastid localization enzyme, formed by a part of the sequence of the N-terminal mature part of a plant gene coding for a plastid localization enzyme, such as described in EP-  
25 A-508,909.

          The plant material may have been, or may subsequently be - further transformed with a polynucleotide comprising a region encoding a protein capable of providing the plant with resistance or tolerance to herbicides, insects, desiccation and/or fungal, bacterial or viral infections, or with a polynucleotide capable of encoding proteins which provide for  
30 improved quality traits such as increased yield, altered starch quality and/or increased nutrient content.

The protein capable of providing for herbicide resistance may be selected from the group consisting of glyphosate oxido-reductase (GOX), 5-enol-pyruvyl-3-phosphoshikimate synthetase (EPSPS), phosphinothricin acetyl transferase (PAT), hydroxyphenyl pyruvate dioxygenase (HPPD), glutathione S transferase (GST), cytochrome P450, Acetyl-CoA carboxylase (ACCase), Acetolactate synthase (ALS), protoporphyrinogen oxidase (PROTOX), dihydropteroate synthase, polyamine transport proteins, superoxide dismutase (SOD), bromoxynil nitrilase, the product of the *tfdA* gene obtainable from *Alcaligenes eutrophus*, and known mutagenised or otherwise modified variants of the said proteins.

As indicated above, the polynucleotide with which the plant material may be transformed may comprise 5' of the protein encoding regions which encode: (i) a peptide which is capable of targeting the translation products of the regions to plastids such as chloroplasts, mitochondria, other organelles or plant cell walls; and/or (ii) non-translated translational enhancing sequences.

The polynucleotide may be codon-optimised, or otherwise altered to enhance at least transcription once it is incorporated into plant material. Thus the polynucleotide used to transform the material may be modified in that mRNA instability encoding motifs and/or fortuitous splice regions may be removed, or plant preferred codons may be used so that expression of the thus modified polynucleotide in a plant yields substantially similar protein having a substantially similar activity/function to that obtained by expression of the unmodified polynucleotide in the organism in which the protein encoding regions of the unmodified polynucleotide are endogenous, with the *proviso* that if - in respect of the herbicide resistance conferring regions - the thus modified polynucleotide comprises plant preferred codons, the degree of identity between the protein encoding regions within the modified polynucleotide and like protein encoding regions endogenously contained within the said plant and encoding substantially the same protein is less than about 70%.

Transformation techniques are well known and include particle mediated biolistic transformation, *Agrobacterium*-mediated transformation, protoplast transformation (optionally in the presence of polyethylene glycols); sonication of plant tissues, cells or protoplasts in a medium comprising the polynucleotide or vector; micro-insertion of the polynucleotide or vector into totipotent plant material (optionally employing the known silicon carbide "whiskers" technique), electroporation and the like.

The invention still further provides morphologically normal fertile (or male sterile) whole plants regenerated from the material mentioned in the paragraph immediately preceding the last and the progeny of such plants, the seed of such plants and progeny, and parts of such plants and progeny. The transformed inventive plants include small grain cereals, oil seed  
5 crops, fibre plants, fruit, vegetables, plantation crops and trees. Particularly preferred such plants include soybean, cotton, tobacco, sugarbeet, oilseed rape, canola, flax, sunflower, potato, tomato, alfalfa, lettuce, maize, wheat, sorghum, rye, bananas, barley, oat, turf grass, forage grass, sugar cane, pea, field bean, rice, pine, poplar, apple, grape, citrus and nut plants.

The transformed plants of the invention have tolerance or resistance to certain  
10 herbicides such as the isoxazoles described especially in French Patent Applications 9506800 and 95 13570 and especially of 4-[4-CF<sub>3</sub>-2-(methylsulphonyl)benzoyl]-5-cyclopropylisoxazole, and especially isoxaflutole, a selective maize herbicide, the diketonitriles such as those described in EP-A-496,630 and EP-A-496,631, in particular 2-cyano-3-cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-CF<sub>3</sub>-phenyl)propane-1,3-dione and 2-cyano-3-  
15 cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-2,3-Cl<sub>2</sub>-phenyl)propane-1,3-dione, and the triketones described in EP-A-625,505 and EP-A-625,508, in particular sulcotrione, mesotrione (BSI-proposed), pyrazolynate and pyrazoxyfen.

The invention further includes a morphologically normal fertile (or male sterile) whole plant resulting from the method of the invention, the progeny of such plants, the seed  
20 of such plants and progeny, and parts of such plants and progeny.

The invention still further provides the use of a polynucleotide comprising a region encoding a phytoene desaturase in the production of plant material which is resistant or tolerant to herbicides which - *in vitro* - inhibit the enzyme 4-HPPD.

The invention still further provides a method of selectively controlling weeds in a  
25 field, the field comprising weeds and crop plants, the method comprising application to the field of a herbicide which - *in vitro* - is capable of inhibiting the enzyme 4-HPPD, characterised in that the plants have been transformed with and express the coding regions of a polynucleotide comprising a sequence encoding a phytoene desaturase.

It is particularly preferred that the phytoene desaturase encoding sequence is that  
30 which is depicted in SEQ ID No. 1, or is complementary to one which when incubated at a temperature of between 55 and 60°C in 0.3 strength citrate buffered saline containing 0.1%

SDS followed by rinsing at the same temperature with 0.3 strength citrate buffered saline containing 0.1% SDS still hybridises with the sequence depicted in SEQ ID No. 1. The herbicide may be selected from the group consisting of mesotrione (BSI-proposed), pyrazolynate and pyrazoxyfen, Balance,<sup>TM</sup> sulcotrione etc. The field may be treated with a pesticide selected from the group consisting of a fungicide, insecticide and nematocide, either prior to or post application to the field of the herbicide.

The invention will now be described by way of the following non-limiting example, figure and the Sequence Listing in which:

SEQ ID No. 1 is the sequence of the phytoene desaturase (dehydrogenase) gene isolated from *Erwinia uredovora*. The person skilled in the art will recognise that any phytoene desaturase gene may be used in the production of plants having resistance/tolerance to the herbicides described above.

SEQ ID No. 2 is the protein encoded by SEQ ID No 1.

SEQ ID No.3 is the polynucleotide sequence encoding the pea rubisco small subunit transit peptide.

SEQ IN No. 4 is the amino acid sequence encoded by SEQ ID No. 3.

Figure 1 is the structure of plasmid pYPEIT4 carrying the *Erwinia uredovora* crtI gene with the transit peptide sequence (depicted as TP) of the pea rubisco small subunit.

## EXAMPLE

### **Production of plants tolerant to herbicides capable of inhibiting the enzyme 4-HPPD *in vitro*.**

The PDS gene (crtI) was cloned from *Erwinia uredovora*, a non-green phytopathogenic bacterial rot, and over-expressed in transgenic tobacco and tomato using a plasmid containing the CaMV 35S promoter and a chloroplast transit peptide (pYPIET4) (Misawa *et al.*, 1993). Homozygous tomato lines over-expressing the crtI gene were obtained as were tobacco plants containing the same construct.

### **Construction of plasmid pYPIET4 carrying the tp-crtI gene**



Recombinant DNA techniques were performed using standard methods. A DNA sequence coding for the transit peptide (TP) in the precursor of the ribulose-1,5-bisphosphate carboxylase (Rubisco) small subunit of pea was isolated from plasmid pSNIF83 (Schreier *et al.*, 1985) as a 204 bp HindIII-SphI fragment, whose SphI site contains the tp processing site.

5 Plasmid pCRT-1 (Fraser *et al.* (1992) J.Biol.Chem 267 19891-19895) carrying the intact phytoene desaturase gene (*crtI*) of *Erwinia uredovora* was digested with BamHI and HindIII, and a 1.57 kb BamHI-HindIII fragment carrying the truncated *crtI* gene was isolated. The above 204 bp HindIII-SphI TP fragment was ligated with a 76bp synthesized fragment which carries the reading frame from the cohesive end for the SphI site containing the *crtI*

10 initiation codon to that of the BamHI site, and with the 1.57 kb BamHI-HindIII fragment. The desired 1.84 kb HindIII fragment carrying the tp-*crtI* chimeric gene was isolated, filled in with Klenow enzyme, and ligated into the SmaI-SacI site of a 10.9 kb fragment removing the  $\beta$ -glucuronidase gene from the binary vector bB/121 (purchased from Clontech laboratories). Thus, the desired plasmid pYPEIT4 was created, shown in Figure 1. The initiation codons

15 for the transit peptide and the intact *CrtI* are underlined. This HindIII fragment carrying the tp-*crtI* gene is surrounded by the CaMV 35S promoter and the NOS terminator of the binary vector pBI121 in order to lead to sufficient expression in the tissues of transgenic tobacco and tomato plants. As a control, plasmid pBICAR4 was constructed which carries an intact *crtI* gene without tp surrounded by the CaMV 35S promoter and the NOS terminator. The

20 plasmid pYPEIT4 was introduced into tobacco and tomato material by known techniques and the material then regenerated into intact plants, again by known techniques.

#### **Tolerance of Tomato Plants transformed with *crtI* gene to Mesotrione and Isoxaflutole**

Homozygous seed of tomato plants cv. Ailsa Craig, derived from 'wild type' (i.e. un-

25 transformed) and plants transformed with the *crtI* gene from *Erwinia uredovora*, (see above) were sown in a peat-based compost in 3 inch pots and transferred to the glasshouse. Plants were grown at 20/16 degrees day/night temperature under a 16 hour photoperiod for approximately 4 weeks prior to post-emergence treatment of four replicates with mesotrione or isoxaflutole (Balance <sup>TM</sup> Herbicide) at the 3 leaf stage. The chemicals were suspended in

30 water and applied, via a track sprayer at a spray volume of 200 litres per hectare, at rates ranging from 1 to 500 grammes active ingredient per hectare (g a.i./ha), as shown in Table 1.

The plants were left to grow for a further 25 days and then assessed visually for herbicidal damage compared to untreated 'control' plants. Typical phytotoxic symptoms observed were extreme chlorosis/bleaching and necrosis of leaves and new growth. The results from this test are given in Table 1 below where the '% Damage/Phytotoxicity' scores represent the  
5 mean of the visual assessment from each of the four treatment replicates.

**Table 1**

Chemical	Rate (g a.i./ha)	% Damage/Phytotoxicity (25 days after treatment)	
		Wild Type (Un-Transformed)	Transformed (crtI )
Mesotrione	1	25	0
	3	29	0
	11	50	9
	33	81	49
Isoxaflutole (Balance™)	1	11	2
	5	15	4
	15	19	6
	50	21	8
	150	34	19
	500	65	31

- As can be seen, plants transformed with the crtI gene which expresses the bacterial PDS from *Erwinia uredovora*, demonstrate elevated tolerance to mesotrione and isoxaflutole compared to wild type, un-transformed tomatoes. For example, 11 g a.i./ha of mesotrione caused 50% phytotoxicity to wild type tomatoes but only 9% injury is observed in the transformed plants. Similarly, wild type plants are significantly more damaged by 500 g a.i./ha of isoxaflutole than those containing the crtI gene.
- The skilled man will recognise that the invention is not limited to that described above. For example, plants other than tomato and tobacco may be transformed with a gene encoding a PDS enzyme, whether derived from a bacterial source or otherwise.

CLAIMS

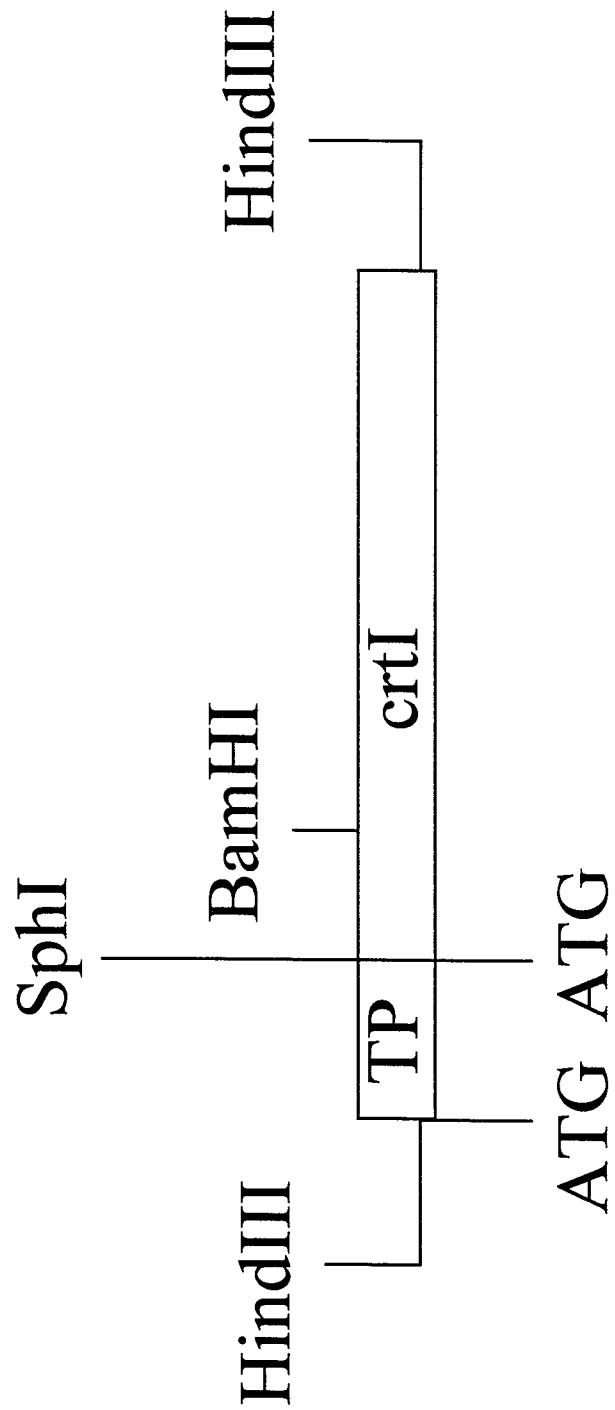
1. A method of making plants which are resistant or tolerant to herbicides which - *in vitro* - inhibit 4-hydroxyphenylpyruvate dioxygenase (4HPPD) comprising the steps  
5 of:
  - (i) transforming plant material with a polynucleotide comprising a region encoding a phytoene desaturase (PDS);
  - (ii) regenerating the thus transformed material into morphologically normal plants and selecting from the population of regenerants those plants which are resistant or  
10 tolerant to herbicides which *in vitro* inhibit 4HPPD.
2. A method according to claim 1, wherein the region comprised by the polynucleotide is the sequence depicted in SEQ ID No. 1, or is a sequence which is complementary to one which when incubated at a temperature of between 55 and 60°C in 0.3 strength  
15 citrate buffered saline containing 0.1% SDS followed by rinsing at the same temperature with 0.3 strength citrate buffered saline containing 0.1% SDS still hybridises with the sequence depicted in SEQ ID No. 1.
3. A method according to claim 1, wherein the phytoene desaturase is of plant origin.  
20
4. A method according to claim 1, wherein the phytoene desaturase is of bacterial origin.
5. A method according to claim 4 wherein the phytoene desaturase is isolatable from  
25 *Erwinia uredovora*.
6. A method according to any one of claims 1 to 5 wherein the polynucleotide further comprises a selectable marker gene.

7. A method according to claim 6 wherein the said selectable marker gene is selected from the group consisting of antibiotic resistance conferring, herbicide resistance conferring, toxin resistance conferring, nutritional markers, visual markers and marker genes used in hormone based selection systems.
- 5
8. A method according to any one of claims 1 to 7, wherein the plant material has been or is further transformed with a polynucleotide comprising a region encoding a protein capable of providing the plant material with resistance or tolerance to herbicides, insects, desiccation and/or fungal, bacterial or viral infections, or with a
- 10 polynucleotide capable of encoding proteins which provide for improved quality traits such as increased yield, altered starch quality and/or increased nutrient content.
9. A method according to any one of claims 1 to 8, wherein the protein encoding sequences within the polynucleotide are bounded by plant operable promoters and
- 15 terminators.
10. A method according to either of claims 8 or 9, wherein the protein capable of providing for herbicide resistance is selected from the group consisting of glyphosate oxido-reductase (GOX), 5-enol-pyruvyl-3-phosphoshikimate synthetase (EPSPS),
- 20 phosphinothricin acetyl transferase (PAT), hydroxyphenyl pyruvate dioxygenase (HPPD), glutathione S transferase (GST), cytochrome P450, Acetyl-CoA carboxylase (ACCase), Acetolactate synthase (ALS), protoporphyrinogen oxidase (PROTOX), dihydropteroate synthase, polyamine transport proteins, superoxide dismutase (SOD), bromoxynil nitrilase, the product of the *tfdA* gene obtainable from
- 25 *Alcaligenes eutrophus*, farnesyl pyrophosphate synthase and known mutagenised or otherwise modified variants of the said proteins.

11. A method according to any one of claims 1 to 10, wherein the protein encoding sequences of the polynucleotide comprise 5' regions which encode: (i) a peptide which is capable of targeting the translation products of the regions to plastids such as chloroplasts, mitochondria, other organelles or plant cell walls; and/or (ii) non-translated translational enhancing sequences.
12. A method according to any one of claims 1 to 11, in which the polynucleotide used to transform the material is modified in that mRNA instability encoding motifs and/or fortuitous splice regions are removed, or plant preferred codons are used so that expression of the thus modified polynucleotide in a plant yields substantially similar protein having a substantially similar activity/function to that obtained by expression of the unmodified polynucleotide in the organism in which the protein encoding regions of the unmodified polynucleotide are endogenous, with the *proviso* that if - in respect of the herbicide resistance conferring regions - the thus modified polynucleotide comprises plant preferred codons, the degree of identity between the protein encoding regions within the modified polynucleotide and like protein encoding regions endogenously contained within the said plant and encoding substantially the same protein is less than about 70%.
13. A method according to any one of claims 1 to 12, in which the 4-HPPD inhibiting herbicide is selected from the group consisting of isoxaflutole, diketonitriles such as 2-cyano-3-cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-CF<sub>3</sub>-phenyl)propane-1,3-dione and 2-cyano-3-cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-2,3Cl<sub>2</sub>phenyl)propane-1,3-dione, triketones such as sulcotrione, and mesotrione (BSI-proposed), pyrazolynate and pyrazoxyfen.
14. A method according to any one of claims 1 to 13, wherein the herbicide is applied post-germination.
15. A morphologically normal fertile (or male sterile) whole plant resulting from the method of any one of claims 1 to 14, the progeny of such plants, the seed of such plants and progeny, and parts of such plants and progeny.

16. A plant according to claim 15 selected from the group consisting of banana, cotton, maize, tomato, vines.
- 5 17. Use of a polynucleotide comprising a region encoding a phytoene desaturase in the production of plant material which is resistant or tolerant to herbicides which - *in vitro* - inhibit the enzyme 4-HPPD.
- 10 18. A method of selectively controlling weeds in a field, the field comprising weeds and crop plants, the method comprising application to the field of a herbicide which - *in vitro* - is capable of inhibiting the enzyme 4-HPPD, characterised in that the plants have been transformed with and express the coding regions of a polynucleotide comprising a sequence encoding a phytoene desaturase.
- 15 19. A method according to claim 18 wherein the polynucleotide is that mentioned in any one of claims 2 to 12.
- 20 20. A method according to either of claims 18 or 19, wherein the said herbicide is selected from the group consisting of, isoxaflutole, diketonitriles such as 2-cyano-3-cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-CF<sub>3</sub>-phenyl)propane-1,3-dione and 2-cyano-3-cyclopropyl-1-(2-SO<sub>2</sub>CH<sub>3</sub>-4-2,3Cl<sub>2</sub>phenyl)propane-1,3-dione, triketones such as sulcotrione, and mesotrione (BSI-proposed), pyrazolynate and pyrazoxyfen.
- 25 21. A method according to any one of claims 18 to 20, wherein the field is treated with a pesticide selected from the group consisting of a fungicide, insecticide and nematocide, either prior to or post application to the field of the herbicide.

1/1

**FIGURE 1**



-1-

## SEQUENCE LISTING

&lt;110&gt; ZENECA LIMITED

<120> METHOD OF PRODUCING PLANTS WHICH ARE TOLERANT OR  
RESISTANT TO HERBICIDES

&lt;130&gt; PPD50336WO

&lt;140&gt;

&lt;141&gt;

&lt;150&gt; 9807818.1

&lt;151&gt; 1998-04-09

&lt;160&gt; 4

&lt;170&gt; PatentIn Ver. 2.0

&lt;210&gt; 1

&lt;211&gt; 1493

&lt;212&gt; DNA

&lt;213&gt; Erwinia uredovora

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (15)..(1493)

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 <213> Erwinia uredovora

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&lt;210&gt; 3

&lt;211&gt; 204

&lt;212&gt; DNA

&lt;213&gt; Pisum sativum

&lt;220&gt;

-5-

&lt;221&gt; CDS

&lt;222&gt; (1)..(204)

&lt;400&gt; 3

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Met Thr Gly Phe Pro Val Lys Lys Val Asn Thr Asp Ile Thr Ser Ile
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aca agc aat ggt gga aga gta aag tgc atg aaa cca act acg gta att 192
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&lt;210&gt; 4

&lt;211&gt; 68

&lt;212&gt; PRT

&lt;213&gt; Pisum sativum

&lt;400&gt; 4

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## INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 99/01059

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/82 A01H5/00 A01H5/10 //C12N15/53

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N A01H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	MISAWA N ET AL: "EXPRESSION OF AN ERWINA PHYTOENE DESATURASE GENE NOT ONLY CONFERS MULTIPLE RESISTANCE TO HERBICIDES INTERFERING WITH CAROTENOID BIOSYNTHESIS BUT ALSO ALTERS XANTHOPHYLL METABOLISM IN TRANSGENIC PLANTS" PLANT JOURNAL, vol. 6, no. 4, 1 January 1994 (1994-01-01), pages 481-489, XP002017203 ISSN: 0960-7412 the whole document	15, 16
A	page 486, column 1, paragraph 2; table 1 ---	
A	WO 97 27285 A (UNIV ARIZONA) 31 July 1997 (1997-07-31) page 6, line 7-24 --- -/--	

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

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"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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"&" document member of the same patent family

Date of the actual completion of the international search

11 August 1999

Date of mailing of the international search report

30/08/1999

Name and mailing address of the ISA

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Fax: (+31-70) 340-3016

Authorized officer

Bilang, J

## C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>BARTA I C ET AL: "BENZOYLCYCLOHEXANEDIONE HERBICIDES ARE STRONG INHIBITORS OF PURIFIED P-HYDROXYPHENYLPYRUVIC ACID DIOXYGENASE OF MAIZE" PESTICIDE SCIENCE, vol. 45, no. 3, 1 November 1995 (1995-11-01), page 286/287 XP000547268 ISSN: 0031-613X the whole document</p> <p>----</p>	
A	<p>SCHULZ A ET AL: "SC-0051, A 2-BENZOYL-CYCLOHEXANE-1,3-DIONE BLEACHING HERBICIDE, IS APOTENT INHIBITOR OF THE ENZYME P-HYDROXYPHENYLPYRUVATE DIOXYGENASE" FEBS LETTERS, vol. 318, no. 2, 1 March 1993 (1993-03-01), pages 162-166, XP002028049 ISSN: 0014-5793 abstract</p> <p>----</p>	
A	<p>BABCZINSKI P ET AL: "SUBSTITUTED TETRAHYDROPYRIMIDINONES. A NEW CLASS OF HERBICIDAL COMPOUNDS INDUCING CHLOROSIS BY INHIBITION OF PHYTOENE DESATURATION" PESTICIDE SCIENCE, vol. 30, no. 3, 1 January 1990 (1990-01-01), pages 339-342, XP000202878 ISSN: 0031-613X the whole document</p> <p>----</p>	
A	<p>MISAWA ET AL.: "E. uredovora carotenoid biosynthesis genes" EMBL DATABASE ACCESSION NUMBER D90087, 6 January 1991 (1991-01-06), XP002111545 the whole document</p> <p>-----</p>	

# INTERNATIONAL SEARCH REPORT

### Information on patent family members

Interr      al Application No

PCT/GB 99/01059

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9727285 A	31-07-1997	AU 1845397 A EP 0877793 A	20-08-1997 18-11-1998
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